

HIGHLY OXYGENATED FLAVONOIDS FROM *MELICOPE TRIPHYLLA*

TING-TING JONG and TIAN-SHUNG WU

Department of Applied Chemistry, Providence College of Arts and Science, Shalu 43309, Taichung Hsien, Taiwan, Republic of China

(Received in revised form 20 June 1988)

Key Word Index—*Melicope triphylla*; Rutaceae; flavonoid; furoquinoline; melicophyllin; anti-platelet aggregation activity.

Abstract—The investigation of the leaves of *Melicope triphylla* resulted in the isolation of two furoquinoline alkaloids, skimmianine, kokusagenine and five known flavones, meliternin, melisimplexin, melibentin, meliternatin, 3,5,8-trimethoxy-6,7; 3',4'-dimethylenedioxyflavone and a new flavone, melicophyllin. The latter was shown to be 3,5,8,3',4'-pentamethoxy-6,7-methylenedioxyflavone from its spectroscopic properties. Three of these flavones were found to possess anti-platelet aggregation activity at 100 µg/ml (*in vitro*).

INTRODUCTION

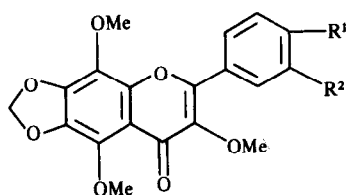
Melicope triphylla, a small evergreen tree, is the only species of *Melicope* native to Taiwan. Furoquinoline alkaloids and polymethoxyflavonoids—including very rare methylenedioxyflavones—have been reported to be the major components in this genus. The antifeedant activity of the leaves extract of this plant [1] and anti-platelet aggregation activity of polymethoxyflavonoids [2] aroused our interest in this plant. Higa *et al.* have reported some flavonoids and furoquinoline alkaloids from this plant [1]. In our study, two sesquiterpene lactones with novel skeleta were obtained from the root bark [3]. From the leaves we have now obtained six polymethoxyflavonoids, each containing one or two methylenedioxy substituents. They are meliternin (2) [4], melisimplexin (3) [5], melibentin (4) [6], meliternatin (5) [7], 3,5,8-trimethoxy-6,7; 3',4'-dimethylenedioxyflavone (6) [8] and a new flavone-melicophyllin (1), as well as two furoquinoline alkaloids, skimmianine (7) and kokusagenine (8). We now report the isolation and structural elucidation of 1 and anti-platelet aggregation activity of these flavonoids.

RESULTS AND DISCUSSION

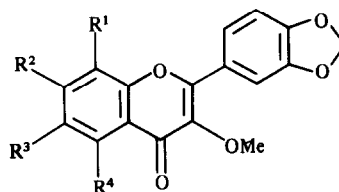
Known compounds, meliternin (2), melisimplexin (3), melibentin (4), meliternatin (5), 3,5,8-trimethoxy-6,7; 3',4'-dimethylenedioxyflavone (6) and skimmianine (7), kokusagenine (8) were characterized based on the spectroscopic analyses and comparisons with the authentic samples.

Melicophyllin (1), mp 175–177°, C₂₁H₂₀O₉ (M⁺, 416) was shown to be a flavonoid from UV absorption bands at λ_{max} 255, 275 and 339 nm, together with IR spectrum showing a peak at 1640 cm⁻¹ (α, β-unsaturated carbonyl). ¹H NMR spectrum of 1 showed five methoxy groups at δ 4.06 (6H, s), 3.96 (6H, s), 3.89 (3H, s) and methylenedioxy protons at δ 6.08 (2H, s). At the downfield region, there is one ABX type protons at δ 7.00 (1H, d, J = 9 Hz, H-5'); 7.79 (1H, d, J = 2 Hz, H-2') and 7.81 (1H, dd, J = 2 and 9 Hz, H-6'), suggesting that 3',4'-positions in ring B are substituted. From the above spectroscopic data, we concluded that melicophyllin (1) is an isomer of melibentin (4), for which there are three possible structures.

From the mass spectrum of 1, through the presence of a peak at m/z 209, the A ring has one methylenedioxy group



- 1 R¹ = R² = OMe
6 R¹, R² = —OCH₂O—



- 2 R¹ = R² = R⁴ = OMe, R³ = H
3 R¹ = H, R² = R³ = R⁴ = OMe
4 R¹ = R² = R³ = R⁴ = OMe
5 R¹ = H, R², R³ = —OCH₂O—, R⁴ = OMe

Table 1. ^1H NMR spectra of flavonoids 1–6.

| | 1 | | 2 | | 3 | | 4 | | 5 | | 6 | |
|-----------------------------|-----------------|------------------------|-----------------|------------------------|-----------------|------------------------|-----------------|------------------------|-----------------|------------------------|-----------------|------------------------|
| | CDCl_3 | C_6D_6 | CDCl_3 | C_6D_6 | CDCl_3 | C_6D_6 | CDCl_3 | C_6D_6 | CDCl_3 | C_6D_6 | CDCl_3 | C_6D_6 |
| 3-OMe | 3.89 | 3.88 | 3.88 | 3.79 | 3.86 | 3.80 | 3.89 | 3.80 | 3.86 | 3.78 | 3.89 | 3.78 |
| 5-OMe | 4.06* | 4.04 | 3.99* | 3.47 | 4.00 | 4.06 | 4.09 | 4.02 | 4.12 | 4.00 | 4.07* | 4.01 |
| 6-OMe (or H) | | | 6.41 | 5.91 | 3.96* | 3.76 | 3.97* | 3.73 | | | | |
| 7-OMe | | | 3.92* | 3.22 | 3.91* | 3.18 | 3.94* | 3.75 | | | | |
| 8-OMe (or H) | 4.05* | 3.73 | 3.99* | 3.70 | 6.73 | 6.23 | 3.99* | 3.62 | | | 4.05* | 3.66 |
| 6,7-O-CH ₂ -O- | 6.08 | 5.08 | | | | | | | 6.04 | 5.05 | 6.08 | 5.03 |
| 2'-H | 7.79 | 7.85 | 7.69 | 7.88 | 7.59 | 7.72 | 7.67 | 7.84 | 7.56 | 7.66 | 7.70 | 7.87 |
| 3',4'-O-CH ₂ -O- | | | 6.05 | 5.23 | 6.06 | 5.26 | 6.07 | 5.26 | 6.04 | 5.25 | 6.06 | 5.23 |
| 3'-OMe | 3.96 | 3.54 | | | | | | | | | | |
| 4'-OMe | 3.96 | 3.36 | | | | | | | | | | |
| 5'-H | 7.00 | 6.58 | 6.94 | 6.68 | 6.93 | 6.70 | 6.95 | 6.68 | 6.90 | 6.67 | 6.94 | 6.67 |
| 6'-H | 7.81 | 7.94 | 7.77 | 7.83 | 7.67 | 7.66 | 7.76 | 7.80 | 7.63 | 7.58 | 7.79 | 7.82 |

Values are in ppm.

*Values with same superscript may be interchanged.

and two methoxy groups and there appeared fragmentation ions at m/z $[\text{M}-1]^+$, and m/z $[\text{M}-19]^+$, indicating the C-3 and C-5 were substituted with methoxy groups [9]. However, the peak at m/z 401 $[\text{M}-15]^+$ is not the base peak; therefore, the presence of a C-6 methoxy group can be excluded [9]. In the ^1H NMR spectrum in C_6D_6 , the shifts observed for the methoxy groups of the A ring excluded the presence of a methoxy group at C-7 [10]. Consequently, the methylenedioxy group is located at C-6, C-7 position and **1** is 3,5,8,3',4'-pentamethoxy-6,7-methylenedioxyflavone.

This structure is further confirmed by the comparison of the ^1H NMR spectrum of **1** with other five known flavones isolated from this plant (Table 1). Thus the chemical shift for the methylenedioxy protons of **1** in C_6D_6 is similar to that of compound (**5**) and (**6**) with 6,7-substituted methylenedioxy groups. Second, the chemical shifts of H-2' and H-6' on ring-B shifted downfield (>0.1 ppm), whenever H-8 was substituted by methoxy group in ring-A both in CDCl_3 and C_6D_6 . This is an effective means to decide if H-8 is substituted by a methoxyl group in the flavonoid series.

The highly oxygenated flavonoids (**2**–**6**) were subjected to both ADP and collagen induced platelet aggregation tests and results shown meliternin (**2**), meliternatin (**5**) and 3,5,8-trimethoxy-3',4'; 6,7-dimethylenedioxyflavone (**6**) at 100 $\mu\text{g}/\text{ml}$ (*in vitro*) were found to possess medium inhibitory activity toward collagen-induced platelet aggregation.

EXPERIMENTAL

UV spectra were run in MeOH and IR spectra as KBr discs. ^1H NMR spectra were run at 100 MHz in CDCl_3 using TMS as internal standard. MS were obtained at 70 eV direct inlet system. Mps: uncorr.

Plant material. *Melicope triphylla* was collected from Orchid Island (Lan-Yu) in Taiwan on September, 1985 and verified by Prof. C. S. Kuoh. The specimen is deposited in the Herbarium of Cheng-Kung University, Taiwan, Republic of China.

Extraction and separation. Air-dried and powdered leaves (645 g) of *Melicope triphylla* were extracted with hot Me_2CO (1 l

$\times 7$). The Me_2CO extract was concd and the ppt was identified as skimmianine (1.2 g). The filtrate was partitioned between CHCl_3 and H_2O . The CHCl_3 layer was extracted with 5% HCl soln. The acidic layer was neutralized with NH_4OH and extracted with CHCl_3 . After concn and chromatography over silica gel (CHCl_3 – Me_2CO 19:1), skimmianine (120 mg), kokusagenine (25 mg) and two unknown compounds were obtained. The CHCl_3 layer after removal of the basic portions was chromatographed on silica gel and eluted exhaustively with CHCl_3 – Me_2CO (19:1) to afford successively, melisimplexin (**3**) (23 mg), melibentin (**4**) (35 mg), 3,5,8-trimethoxy-3',4'; 6,7-dimethylenedioxyflavone (**6**) (10 mg), meliternatin (**5**) (23 mg), melicophyllin (**1**) (20 mg), and meliternin (**2**) (28 mg).

Melicophyllin (1). Colourless crystals. mp 175–177° (Me_2CO). UV λ_{max} nm: 255, 275, 339; IR ν_{max} cm^{-1} 1640, 1620, 1600. MS m/z (%): 416 (M^+ , 100), 415 (66), 397 (22), 387 (25), 385 (25), 383 (20), 373 (19), 355 (18), 209 (20), 194 (22), 165 (30), 149 (30).

Acknowledgements—The authors wish to thank the National Science Council of the Republic of China (NSC76-0208-M126-01) for financial support. Biological tests were carried out at the Brion research laboratory, Taipei, Taiwan.

REFERENCES

- Higa, M., Miyagi, Y., Yogi, S. and Hokama, K. (1987) *Yakugaku Zasshi*, **107**, 954.
- Chen, Y. P., Chen, C. C., Cheng, H. T. and Hsu, H. Y. (1986), *Asia-Pacific Symposium on Natural Product Chemistry*, p. 25, Taiwan.
- Wu, T.-S., Jong, T.-T., Ju, W.-M., McPhail, A. T., McPhail, D. R. and Lee, K.-H. (1988) *J. Chem. Soc. Chem. Comm.* 956.
- Briggs, L. H. and Locker, R. H. (1949) *J. Chem. Soc.*, 2157.
- Briggs, L. H. and Locker, R. H. (1950) *J. Chem. Soc.*, 2376.
- Ritchie, E. and Taylor, W. C. (1965) *Aust. J. Chem.*, **18**, 2021.
- Briggs, L. H. and Locker, R. H. (1951) *J. Chem. Soc.*, 3131.
- Fauvel, M. T., Gleye, J., Moulis, C., Blasco, F. and Stanislas, E. (1981) *Phytochemistry* **20**, 2059.
- Kingston, D. G. I. (1971) *Tetrahedron*, **27**, 2691.
- Wilson, R. B., Bowie, J. H. and Williams, D. H. (1968) *Tetrahedron*, **24**, 1407.